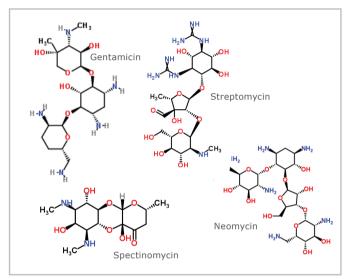
# Extraction and Clean-up of Aminoglycoside Antibiotics from Chicken Meat Using EVOLUTE° WCX Solid Phase Extraction Columns with Analysis by LC-MS/MS

*Jessica Lance<sup>1</sup>, Gloria Gaucin<sup>1</sup>, Carrie Snyder<sup>1</sup>, B. J. Bench<sup>1</sup>, Frank Kero<sup>2</sup>, Victor Vandell<sup>2</sup>, Martin Cherrier<sup>2</sup>, Elena Gairloch<sup>2</sup>* 

1. Tyson Foods, Inc. Springdale Corporate Laboratory, Springdale, AR 72762, USA

2. Biotage, 10430 Harris Oaks Blvd., Suite C, Charlotte, NC USA 28269, USA



## Introduction

This application note describes a polymer based-based weak exchange mixed-mode SPE protocol for the extraction of the aminoglycoside antibiotics gentamicin, streptomycin, spectinomycin and neomycin from chicken meat prior to LC-MS/MS analysis.

The utility of aminoglycoside antimicrobial additives and foods has recently been challenged due to the effects of bioaccumulation in an animal host. The persistence of these selected residues may result in population resistance to antibiotic treatment and as a consequence, has been considered an issue of public health.

This application note describes an approach to extraction of these analytes from chicken meat, with optimized analyte recovery, minimal ion suppression, and acceptable method precision.

Figure 1. Analyte structures

## Analytes

Gentamicin, Streptomicin, Spectinomycin, Neomycin

# Sample Preparation Procedure

Format: EVOLUTE° WCX 500 mg/6 mL solid phase extraction columns, p/n 612-0050-C

#### Sample Pre-treatment:

Weigh 5 g of chicken meat into a 50 mL conical tube.

Spike with internal standard as required.

Add 20 mL of extraction solution (10 mM  $NH_4OAc$ , 0.4 mM EDTA, 0.5% NaCl and 2% trichloroacetic acid in water), vortex for 1 min and shake for 10 mins. After shaking, centrifuge for 10 mins at 4000rpm, and deacant the supernatant into a clean tube taking care not to transfer any of the tissue.

Repeat the extraction procedure and combine the two extracts from each sample. Filter using a glass fibre filter.

Adjust the sample pH to 6.5+/-0.25 using 30% NaOH, 1 N NaOH and 1 N HCl.



# Solid Phase Extraction

Plate Conditioning:	Condition each column with methanol (10 mL)
Plate Equilibration:	Equilibrate each column with water (10 mL)
Sample loading:	Load pre-treated sample (40 mL) using vacuum or positive pressure
Interference Wash :	Elute interferences with water adjusted to pH 6.5 (10 mL)
Elution:	Elute analytes with 25% formic acid in water (6 mL )
Post Extraction:	Add HPLC grade water to the extract to give a total volume of 10 mL, and filter through a 0.2 $\mu m$ disc

# **UPLC** Conditions

Instrument:	Waters Acquity UPLC
Column:	Waters Acquity UPLC BEH Amide 1.7 $\mu$ m 2.1 x 50mm
Mobile Phase:	<b>A:</b> 20 mM heptafluorobutyric acid (HFBA) in H2O/acetonitrile (95/5, v/v) <b>B:</b> 20 mM HFBA in acetonitrile
Flow Rate:	0.2 mL/min

#### Table 1. Gradient Conditions

Flow rate (mL/min)	% A	% B
0.2	100	0
0.2	80	20
0.2	80	20
0.2	60	40
0.2	10	90
0.2	10	90
0.2	100	0
0.2	100	0
0	100	0
	(mL/min) 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2 0.2	(mL/min) A   0.2 100   0.2 80   0.2 60   0.2 10   0.2 10   0.2 10   0.2 100   0.2 100   0.2 100

Injection Volume:	7.5 μL
Sample Temperature:	10 °C
Column Temperature	40 °C

# Mass Spectrometry Conditions

Instrument:	Xevo TQD Triple Quad Mass Spec equipped with an electrospray ionization source operated in positive ion mode. The compound selective MRM transitions are detailed in <b>Table 3</b> .
Desolvation Temperature:	350 °C
Ion Source Temperature:	150 °C
Collision Cell Pressure:	3.4 x 10 <sup>.3</sup> mbar

Positive ions acquired in the multiple reaction monitoring (MRM) mode:



Analyte	MRM Transition	Cone Voltage (V)	Collision Energy (eV)
C1 Gentamicin	479>157,160,322	40	25
C1a Gentamicin	450>112,160,322	35	25
C2+C2a Gentamicin	464>160,163,322	35	20
Streptomycin	582>176,246,263	70	32
Spectinomycin	351>98,140,333	40	20
Neomycin	615>160,163,293	52	30

Table 2. MRM transitions for selected analytes

### Results

A summary of the performance for this method is given in Table 3. The observed linearity for each analyte over the concentration range of interest was  $r^2 > 0.990$ . The reference range defined by USDA-FSIS guidelines was 100-500 ppb.

Table 3. Method performance criteria

Analyte	Linearity	LOQ (ppb)	LOQ spec
Gentamicin	0.9915	33	100
Streptomycin	0.9986	382	500
Spectinomycin	0.9959	75	100
Neomycin	0.9993	361	500

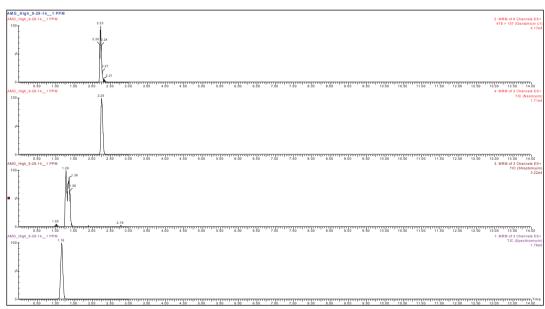


Figure 2. Representative chromatogram of a fortified meat specimen processed using EVOLUTE\* WCX

#### Recovery

Relative recoveries of the selected analyte from fortified specimens were determined at 3 concentration levels. The results are given in **Figure 3**.

Streptomycin and neomycin: Level 1 =400 ppb; Level 2 = 1000 ppb; Level 3 = 10000 ppb

Gentamicin and spectinomycin: Level 1 =50 ppb; Level 2 = 500 ppb; Level 3 = 1000 ppb



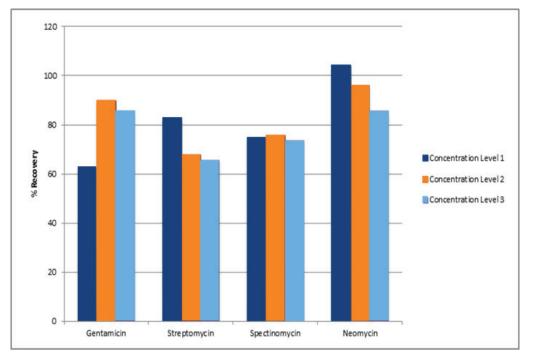


Figure 3. Relative recovery from fortified samples

## Conclusions

This method, utilizing EVOLUTE WCX SPE columns was demonstrated as a viable option for residue measurements over a relevant concentration range in food safety laboratory applications.

## Additional Notes

- 1. Buffer Preparation. Aminoglycoside extraction solvent mixture (10mM NH<sub>4</sub>OAc, 0.4 mM EDTA, 0.5% NaCl and 2% TCA in water): Add 1.54 g of  $NH_4OAc$  to 1.95 L of water. Adjust the pH of the solution to 4.0 with 1N HCl and/or 1 N NaOH. Add 0.3 g Na<sub>2</sub>EDTA.2H<sub>2</sub>O, 10 g of NaCl and 40 g TCA. Mix to ensure the salts dissolve and adjust final volume to 2 L with pure water.
- 2. Processing Conditions. Samples were loaded on the SPE cartridge at a flow rate of 2 mL/min.

## **Ordering Information**

Part Number	Description	Quantity
612-0050-C	EVOLUTE <sup>®</sup> WCX 500 mg/6 mL SPE columns	30
PPM-48	Biotage® Positive Pressure Manifold 48 Position	1

EUROPE	NORTH & LATIN AMERICA	JAPAN
Main Office: +46 18 565900	Main Office: +1 704 654 4900	Tel: +81 3 5627 3123
Toll Free: +800 18 565710	Toll Free: +1 800 446 4752	Fax: +81 3 5627 3121
Fax: +46 18 591922	Fax: +1 704 654 4917	jp_order@biotage.com
Order Tel: +46 18 565710	Order Tel: +1 704 654 4900	jp-1-pointsupport@biotage.com
Order Fax: +46 18 565705	Order Fax: +1 434 296 8217	
order@biotage.com	ordermailbox@biotage.com	
Support Tel: +46 18 56 59 11	Support Tel: +1 800 446 4752	
Support Fax: + 46 18 56 57 11	Outside US: +1 704 654 4900	
eu-1-pointsupport@biotage.com	us-1-pointsupport@biotage.com	

#### APAN Tel: +81 3 5627 3123 ax: +81 3 5627 3121 p\_order@biotage.com

#### CHINA

Tel: +86 21 2898 6655 Fax: +86 21 2898 6153 cn\_order@biotage.com cn-1-pointsupport@biotage.com

Biotage

To locate a distributor, please visit our website at www.biotage.com

#### Part Number: AN833

© 2015 Blotage. All rights reserved. No material may be reproduced or published without the written permission of Biotage. Information in this document is subject to change without notice and does not represent any commitment from Biotage. E&OE. Product and company names mentioned herein may be trademarks or registered trademarks and/or service marks of their respective owners, and are used only for explanation and to the owners' benefit, without intent to infringe. For more information visit www.biotage.com